

# Hypothermic Storage of Amniotic Membranes Conserves Structure, Composition, and Responses *In Vitro* through the Retention of Native Characteristics

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## INTRODUCTION

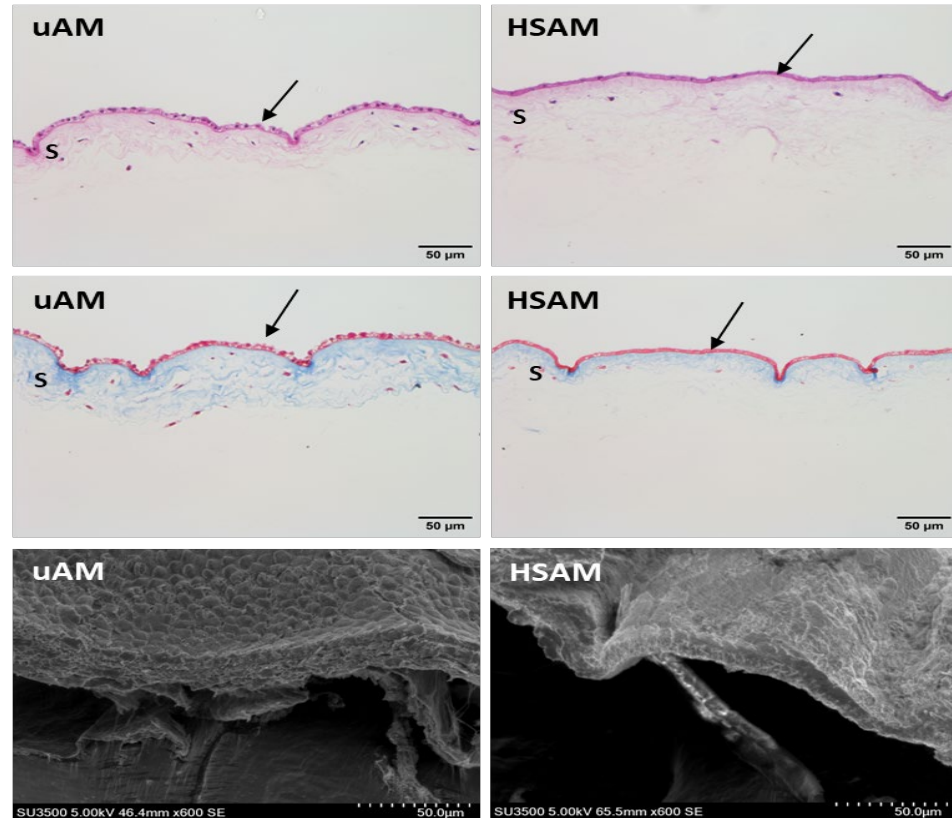
Human placental tissues have an extensive clinical history, while preclinical studies have established that these tissues retain the extracellular matrix (ECM) and serve as protective barriers. A propriety hypothermic storage processing technique has been developed to preserve the native characteristics of amniotic membranes (HSAM\*). In this study, it was hypothesized that the gentle HSAM process would result in the retention of characteristics found in native, unprocessed amniotic membranes (uAM).

\*Affinity®, Organogenesis, Canton, MA

## METHODS

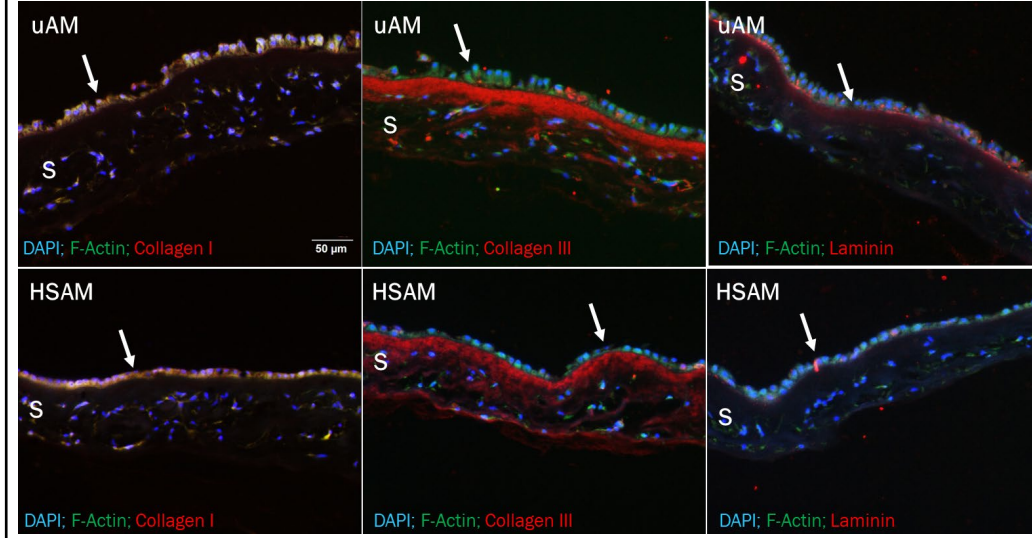
- Donor-matched amniotic membranes were processed into either HSAM, which was stored at 1-10°C for up to 42 days before use, or uAM, which was used within 24 hours.
- Amniotic membranes were characterized using histology, scanning electron microscopy, and immunofluorescence.
- Durability was evaluated using an *in vitro* simulated wound fluid model.
- To evaluate retention of barrier properties, an *in vitro* human dermal fibroblast model was used to assess cell attachment via histology, immunofluorescence staining, and scanning electron microscopy.

## HSAM MAINTAINS UNPROCESSED STRUCTURAL CHARACTERISTICS



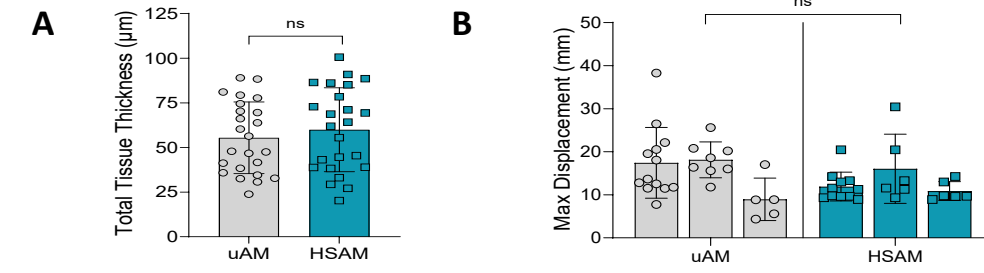
**Figure 1. Structural assessment of uAM and HSAM.** ECM structure was evaluated using H&E and Masson's Trichrome. Imaging revealed maintenance of tissue architecture between uAM and HSAM. Arrow: epithelial layer; S: stromal layer.

## HSAM MAINTAINS UNPROCESSED STRUCTURAL CHARACTERISTICS

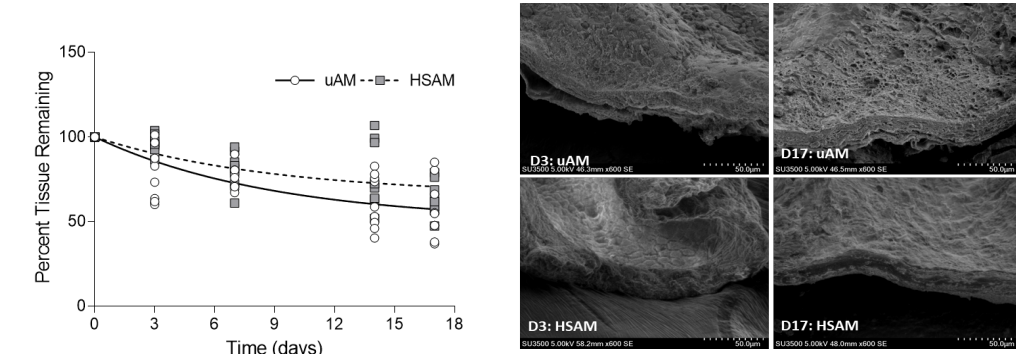


**Figure 2. Structural assessment of uAM and HSAM.** ECM structure was further assessed using immunofluorescence staining including collagen I, collagen III, and laminin. Arrow: epithelial layer; S: stromal layer.

## HYPOTHERMIC STORAGE RETAINS UNPROCESSED TISSUE INTEGRITY

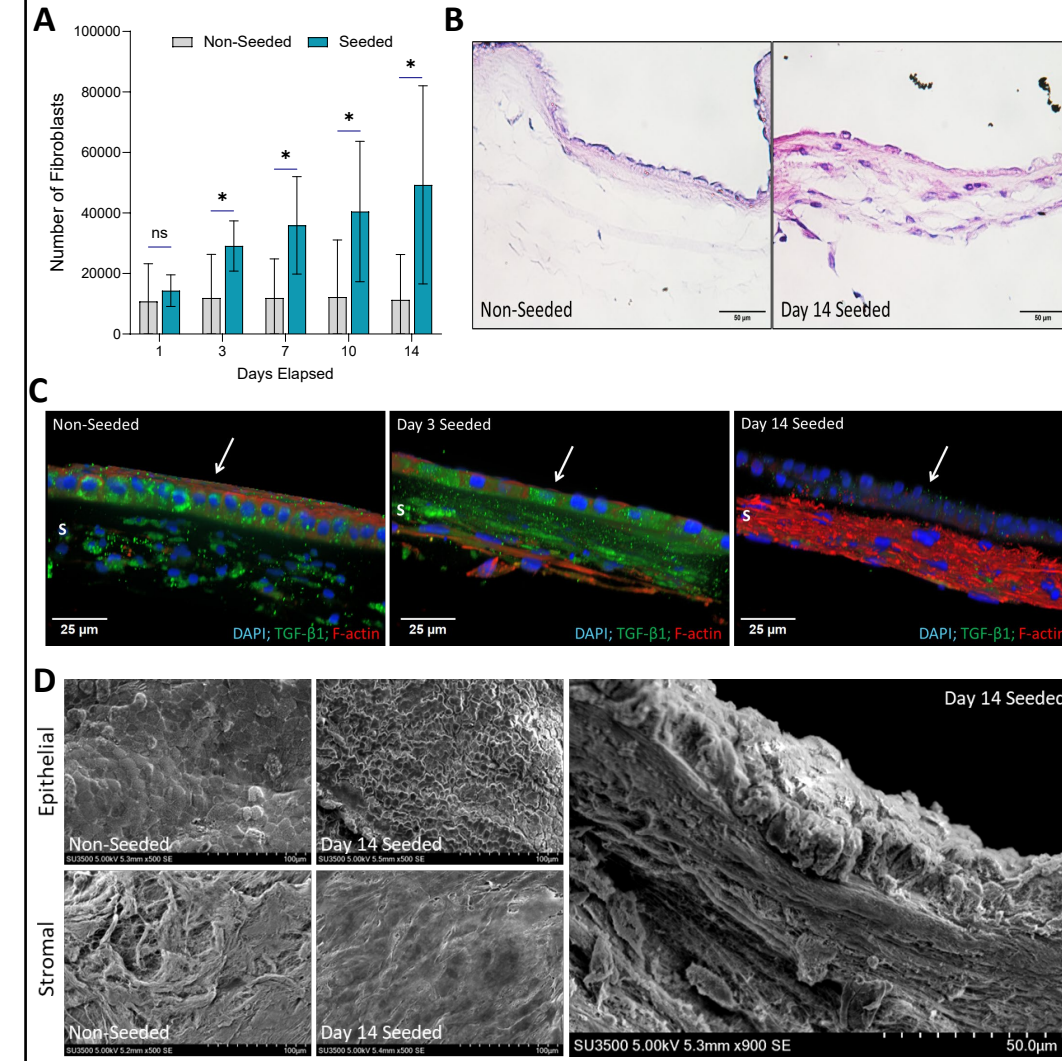


**Figure 3. Characterization of unprocessed and hypothermically stored membranes.** Retention of (A) tissue thickness and (B) tensile testing was evaluated in donor-matched uAM and HSAM. Average  $\pm$  standard deviation reported; ns: not significant.



**Figure 4. Durability of tissues following exposure to SWF *in vitro*.** Rate of degradation and changes in ECM structure were evaluated over 17 days of *in vitro* degradation in SWF. Both uAM and HSAM retained over 50% of their initial tissue weight over the 17-day study, with no significant differences in the rate of degradation or tissue structure.

## HSAM SUPPORTS BARRIER PROPERTIES



**Figure 4. *In vitro* barrier properties.** Fibroblasts were evaluated over 14 days as assessed by (A) metabolic activity, (B) H&E, (C) immunofluorescence staining, and (D) scanning electron microscopy. Average  $\pm$  standard deviation reported; \* $p \leq 0.05$ . Blue: nuclei; green: TGF- $\beta$ 1; red: f-actin (phalloidin); arrow: epithelial layer; S: stromal layer, ns: not significant.

## CONCLUSIONS

- HSAM is a minimally manipulated, protective barrier that retained the characteristics of native, unprocessed amniotic membranes.
- Tissue thickness, tensile strength, and *in vitro* degradation rates were comparable between unprocessed and hypothermically stored amniotic tissues.
- HSAM served as a protective barrier by supporting fibroblasts over 14 days of culture.
- Overall, all facets were conserved with HSAM, due to retaining the properties of uAM. These results highlight the hypothermic processing techniques as an effective method for processing and preserving amniotic membranes.