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## ABSTRACT

**Background:** Nanocrystalline silver wound products have been used for ~30 years. Although they have demonstrated strong biological activity, they are slow and costly to manufacture, both in capital and operating costs, resulting in limited availability. A next-generation process for synthesis of nanostructured silver thin films was developed to address these issues. The study's purpose was to examine the impact of modulating variables in this unique process on the resultant chemistry and antimicrobial activity.

**Methods:** Nanostructured silver was deposited on high density polyethylene (HDPE) by DC magnetron sputtering (99.99% Ag, 12.7x51.4cm target at 15°C, 10 cm working distance). Variables including:

- Ar/O<sub>2</sub> ratio (0%-4.5% O<sub>2</sub>, remainder Ar)
- Water injection rate (0-51.6 μL/min)
- Water injection method (see Results section for details)
- Sputtering power (0.5-1.8A at 380V)
- Run length (15-30min)
- Static or moving HDPE (0-2.0 cm/min), and
- Working gas pressure (13-60 mTorr)

were systematically altered, and the impact on total silver deposited, percent ammonium hydroxide-soluble (active/AHS) silver, and antimicrobial activity was tested. Total silver per unit surface area of dressing was determined by digesting 1"x1" pieces in 20 mL of 50% HNO<sub>3</sub> in distilled H<sub>2</sub>O for 20min. AHS silver was determined by digesting 1"x1" pieces in 20 mL of NH<sub>4</sub>OH for 10min. Both digestions were diluted and analysed by atomic absorption spectrometry (AAS) or inductively coupled plasma mass spectrometry (ICP-MS). Antimicrobial activity was tested via log reductions: A 16h *P. aeruginosa* culture was inoculated into 50 mL calf serum (a wound fluid model) and incubated for 16h to produce a 1x10<sup>9</sup> CFU (colony forming unit) inoculum. Dressings prepared from two silver-coated pieces of HDPE with a piece of gauze in between (2.5x2.5cm) received 200 μL inoculum and were incubated for 1h at 37°C. They were then placed in a sodium thioglycolate/saline/Tween-80 (STS) solution (to inactivate the silver and recover microorganisms), vortexed, serially diluted, and spot plated. CFUs were counted after ~24h incubation at 37°C.

**Results:** Direct injection of H<sub>2</sub>O into the sputtering chamber, and injection of H<sub>2</sub>O into Ar in the absence of O<sub>2</sub>, did not result in sufficient AHS silver or bactericidal activity (log reductions ≤2). However, addition of H<sub>2</sub>O, via approaches listed in the Results section, to appropriate Ar:O<sub>2</sub> blends synergistically increased %AHS silver and bactericidal activity, in some cases generating log reductions >8.3 (total kill). This allows for production at higher power levels (increasing production rate), with lower vacuum requirements, decreasing both capital and operating expenses.

**Conclusion:** Under appropriate conditions, dressings with strong consistent antimicrobial activity can be manufactured at much lower costs, and significantly increased speed, relative to currently available nanocrystalline silver dressings.

## BACKGROUND

- >10M patients globally have chronic wounds, with <50% of wounds healing by 12 weeks
- Chronic wounds are on the rise due to aging populations and comorbidities (e.g., diabetes)
- The cost to heal a chronic wound is ~10K-\$34K USD/wound
- Diabetic foot ulcers have a 5y mortality rate of ~30%, rising to ~50% in the event of amputation
- The wound care market is projected at \$30B USD/y by 2030
  - ~\$16B USD/y attributed to advanced wound healing products

### References:

Jarbrink, Syst Rev 2016;5:152. Denny, Healthcare Quarterly 2014;17:7-10.  
<https://www.marketsandmarkets.com/Market-Reports/wound-care-market-371.html>  
 Fife et al. Adv Wound Care 2017;7:77-94. Sen, Adv Wound Care 2019;8:39-48.

## MATERIALS AND METHODS - MANUFACTURING

**Sputtering Machine – Closed**

Control Panel  
Sputtering Chamber  
Vacuum Pumps

**Sputtering Machine – Open**

Silver Target  
Working Gas Inlets  
Substrate To Be Coated  
Cooling Lines

**Final Product**

Absorbent rayon/polyester non-woven gauze  
Ultrasonic weld

➤ Silver thin films deposited on HDPE in extreme sputtering machine with various modifications to support water injection

➤ Variables systematically altered

**Dressing Laminator and Splitter**

Control Panel  
Individual Rolls  
Ultrasonic Welding and Cutting  
Tri-Layer Dressing Rolls

➤ 2 layers of thin film are assembled with absorbent gauze layer in between and cut to size to create final product

## MATERIALS AND METHODS - TESTING

**Total Silver Analysis**  
1"x1" piece of thin film

20 mL nitric acid (65% solution diluted 1:2 in dH<sub>2</sub>O)

20 min at 100°C; remove dressing

Cool; 1:2 dilution in dH<sub>2</sub>O

**AHS Silver Analysis**  
1"x1" piece of thin film

20 mL of 20 g/L ammonium hydroxide (30%)

5 min at RT; remove dressing

1:5 dilution in dH<sub>2</sub>O

ICP-MS  
Computer Control  
Sample Rack  
Vacuum Pump

Select isolated *P. aeruginosa* colony (MHA)

Incubate (16h, 37°C, 50 mL TSB) → 1 mL → A

Incubate (16h, 37°C, 50 mL calf serum) → 0.2 mL → B

10 g weight

Sterile plastic Test dressing (1"x1")

Incubate (1h, 37°C)

3 mL STS → 1 mL → 9 mL PBS → ... → 20 μL → x3

Incubate (16-24h, 37°C, MHA)

Log<sub>10</sub> Reduction = Log<sub>10</sub>(CFU)<sub>inoculum</sub> - Log<sub>10</sub>(CFU)<sub>recovered</sub>

## RESULTS AND DISCUSSION, CONTINUED

Run Length (min)	H <sub>2</sub> O Injection Method*	% O <sub>2</sub> **	H <sub>2</sub> O Injection Rate (μL/min)	Web Speed (cm/min)	Current (A)	Working Gas Pressure (mTorr)	Total Ag (mg/in <sup>2</sup> )	AHS Ag (%)	Log Reduction
15	1	N/A	35	0	1.5	N/A	1.01	21.7	<2
30	None	0	0	1.5	1.5	40	2.52	3.4	2.1
30	None	0	0	1.5	1.8	40	5.04	3.3	2.8
30	None	1	0	1.5	1.5	40	2.83	9.4	4.9
30	None	2	0	1.5	1.5	40	2.32	21.3	4.9
30	None	3	0	1.5	1.5	40	2.93	22.3	5.0
30	None	4	0	1.5	1.5	40	2.64	26.4	5.0
30	2	0	2	1.5	1.8	40	4.96	2.1	2.7
30	2	0	4	1.5	1.8	40	4.80	2.1	2.1
30	2	0	8	1.5	1.8	40	4.32	3.5	2.1
30	2	0	9	1.5	1.5	40	4.32	3.1	<2
30	2	0	9	1.5	1.5	40	3.21	1.7	2.4
30	2	0	20	1.5	1.5	40	4.16	3.6	<2
30	2	0	33	1.5	1.5	40	4.80	2.1	<2
30	2	1	9	1.5	1.5	40	2.47	32.8	6.2
30	2	2	1	0	1.5	40	4.70	24.2	>8.3
30	2	2	1	1.5	1.5	40	4.70	24.2	>7
30	2	2	3	0	1.5	40	6.86	23.6	>8.3
30	2	2	3	1.5	1.5	40	6.86	23.6	>7
30	2	2	5	0	1.5	40	5.66	22.4	>8.3
30	2	2	5	1.5	1.5	40	5.66	22.4	>7
30	2	2	7	0	1.5	40	4.03	26.0	>8.3
30	2	2	7	1.5	1.5	40	4.03	26.0	>7
30	2	2	9	0	1.5	40	6.67	15.7	>8.3
30	2	2	9	1.5	1.5	40	6.67	19.3	>7
30	2	2	9	1.5	1.5	40	3.29	39.2	6.0
30	2	3	9	1.5	1.5	40	2.74	63.2	6.3
30	2	4	9	1.5	1.5	40	2.99	74.0	6.6
30	2	4.25	6	0	0.5	13	NR	35	3.2
30	2	4.25	6	0	0.9	13	NR	35	4.1
30	2	4.25	6	0	1.8	40	NR	35	5.5
30	2	4.25	6	0	1.8	50	NR	48	4.2
30	2	4.25	9	0	1.8	40	3.46	48.2	7
30	3	1	N/A	0	1.5	40	5.14	41.0	6.9
30	3	1	N/A	0	1.5	40	5.32	45.7	>6
30	4	2	9.1	0	1.5	40	3.20	26.0	7.5
30	4	2	51.6	0	1.5	40	4.11	17.3	7.5

**Results/Discussion**

- Sputtering using H<sub>2</sub>O with various addition methods in absence of O<sub>2</sub> did not result in antimicrobial activity
- Sputtering in increasing O<sub>2</sub> in absence of H<sub>2</sub>O improved antimicrobial activity over H<sub>2</sub>O alone (not total kill)
- Synergistic effect seen when oxygen and H<sub>2</sub>O combined during sputtering process
  - Allows for sputtering at higher power and rate, lower O<sub>2</sub> concentration
- Adding H<sub>2</sub>O upstream of the MFC with various injection rates resulted in total kill at 2% O<sub>2</sub>
- Bubbling Ar through heated water prior to entering the sputtering machine, and adding water via microvalve downstream of the MFC prior to the sputtering chamber, also resulting in highly microbicidal dressings

**Conclusions**

- Dressings with strong consistent antimicrobial activity can be manufactured at much lower costs and significantly increased speed, relative to currently available nanocrystalline silver dressings

**Future Work**

- Continued examination of effect of variables on activity, to fine tune process
- Expansion of *in vitro* testing to demonstrate broad antimicrobial activity and longevity
- *In vitro* and *in vivo* biocompatibility and efficacy testing
- Establish quality systems for regulatory compliance
- Sterilization and stability testing

\* 1 = Water added directly to chamber via microvalve (no Ar or O<sub>2</sub> gas)  
 2 = Water added to Ar upstream of mass flow controller (MFC) via syringe pump with piping heated to 50°C  
 3 = Ar bubbled through heated stainless steel column containing 1400 (1<sup>st</sup> run) or 1500 (2<sup>nd</sup> run) mL H<sub>2</sub>O at 88°C  
 4 = Water added via microvalve downstream of MFC, prior to sputtering chamber, at 21°C

\*\* Remainder Ar.  
 N/A = Not applicable.  
 NR = Not recorded.

Green highlighted data = Log reduction >4 (acceptable by FDA standards). Values with ">" indicates total kill.  
 Blue highlighted data = Log reduction between 3-4 (>3 indicates biocidal)  
 White highlighted data = Log reduction between 2-3 (not biocidal, some measured activity)  
 Grey highlighted data = Log reduction <2 (no activity measured)

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